

# **Comprehensive Evaluation of the ForenSeq<sup>™</sup> DNA Signature Prep Kit** on the MiSeq FGx<sup>™</sup> Next-Generation Sequencing System

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### Introduction

#### Capillary electrophoresis (CE) is the current standard for human identification in forensic DNA laboratories. CE uses fragment analysis to analyze short tandem repeats (STRs). These are the mainstay markers for human identification due to their highly polymorphic and discriminatory power among individuals.

Next-Generation Sequencing (NGS) has grown in popularity in recent years. This is due to its enhanced capability to multiplex and sequence hundreds of forensically relevant markers including STRs and single nucleotide polymorphisms (SNPs) Sequencing of individual STR fragments make it possible to distinguish same-sized alleles that have internal sequence variations and can also aid in linking stutter to its parent allele based on the sequence motif. The ForenSeq<sup>™</sup> DNA Signature Prep Kit for the MiSeq FGx<sup>™</sup> NGS system is an assay that targets 27 autosomal STRs, 24 Y-STRs, 7 X-STRs and 94 identity SNPs, and can include 22 phenotypic SNPs, and 56 biogeographical ancestry SNPs through an additional primer set (Primer Set B). This study aimed to evaluate the performance of the ForenSeq<sup>™</sup> DNA Signature Prep

Kit (Primer Set B) in single source and mixture analysis. In addition, a concordance study was be performed between NGS and CE platforms.





M1 = Male 1, F1 = Female 1.



TH01, and 8 at TPOX.



Figure 1A: D8S1179, TH01, and TPOX loci for the M1:F1 1:20 sample run on the MiSeq FGx<sup>™</sup>.

Figure 1B: D8S1179, TH01, and TPOX loci for the M1:F1 1:20 sample run on the 3500 genetic analyzer. M1 = Male 1, F1 = Female 1. Major contributor should show up as 10, 12 at D8S1179, 7 9.3 at TH01, and 9, 11 at TPOX. Minor contributor should be seen as 12, 13 at D8S1179, 9.3 at





Figure 2B: D8S1179, TH01, and TPOX loci for the F1:M1:M2 10:5:1 sample run on the 3500 genetic analyzer. M1 = Male 1, M2 = Male 2, F1 = Female 1. Major contributor should show up as 10, 12 at D8S1179, 7, 7, 9.3 at TH01, and 9, 11 at TPOX. Minor contributors should be seen as 12, 13 and 10, 13 in D8S1179, 9.3 and 6, 10 in TH01, and 8 and 8, 11 in TPOX. **Table 2.** Genotype of donor(s) from two-person and three-person mixtures at D8S1179, TH01, and TPOX M1 = Male 1 M2 = Male 2 F1 = Female 1

-	111 - 1110 1, 112 - 1110 2, 11 - 101110 1.								
	<b>Mixture Ratio</b>	Locus	M1		<b>F1</b>		<b>M2</b>		
		D8S1179	12	13	10	12			
	M1:F1 1:20	<b>TH01</b>	9.3		7	9.3			
		ТРОХ	8		9	11			
	F1:M1:M2 10:5:1	D8S1179	12	13	10	12	10	13	
		<b>TH01</b>	9.3		7	9.3	6	7	
		TPOX	8		9	11	8	11	

### **Discussion and Conclusions**

In this initial mixture evaluation, the number of alleles detected in single and mixture source samples run on the 3500 Genetic Analyzer and the MiSeq FGx<sup>TM</sup> NGS system were counted and reported. In particular, a full concordance of STR allele call was observed for the major and minor DNA contributors of the whole mixture samples tested between the CE and NGS platform. In addition, all alleles of the minor DNA contributors falling into stutter range position of the parental alleles were confirmed by sequencing for the two-, three- and four person DNA mixture series. This preliminary study showed full concordance of genotypes between CE and NGS platforms for single source and mixture samples. Further analyses are ongoing to evaluate the sensitivity of the ForenSeq<sup>™</sup> DNA Signature Prep Kit and the detection of isoalleles. This may assist with mixture deconvolution, enabling the inclusion or exclusion of reference profiles from mixture samples.

## References

3500XL genetic analyzer for protein quality analysis. Thermo Fisher Scientific - US. (n.d.). Retrieved February 14, 2022 <sup>2</sup>Churchill, Jennifer D., Sarah E. Schmedes, Jonathan L. King, and Bruce Budowle. 2016. "Evaluation of the Illumina® Beta Version ForenSeq<sup>™</sup> DNA Signature Prep Kit for Use in Genetic Profiling." Forensic Science International: Genetics 20 (January): 20–29. <sup>3</sup>Ez1 advanced XL. QIAGEN. (n.d.). Retrieved February 15, 2022 <sup>4</sup>Forenseq DNA Signature Prep Kit. Verogen. (n.d.). Retrieved February 14, 2022 <sup>5</sup>Jäger, Anne C., Michelle L. Alvarez, Carey P. Davis, Ernesto Guzmán, Yonmee Han, Lisa Way, Paulina Walichiewicz, et al. "Developmental Validation of the MiSeq FGx Forensic Genomics System for Targeted Next Generation Sequencing in Forensic DNA Casework and Database Laboratories." Forensic Science International: Genetics 28 (May 2017): 52–70 <sup>6</sup>Köcher, Steffi, Petra Müller, Burkhard Berger, Martin Bodner, Walther Parson, Lutz Roewer, and Sascha Willuweit. "Inter-Laboratory Validation Study of the ForenSeq<sup>™</sup> DNA Signature Prep Kit." Forensic Science International: Genetics36 (September 2018): 77–85. <sup>7</sup>Miseq FGX sequencing system. Verogen. (n.d.). Retrieved February 14, 2022 <sup>8</sup>Xavier, Catarina, and Walther Parson. 2017. "Evaluation of the Illumina ForenSeq<sup>™</sup> DNA Signature Prep Kit -- MPS Forensic Application for the MiSeq FGx<sup>™</sup> Benchtop Sequencer." Forensic Science International: Genetics 28 (May): 188–94.

